

6.4 kb

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Purpose: To repeat & optimize preliminarily 6.4 kb.

Tried mini-prep DAPI as a control

Will try 2 dif. cycling conditions 3 step as well as 2 step.

Colonies will be lysed in 2 different ways 1. in PK (single
2. in H₂O colony
improved will also be included again. buffer

conditions: - since 200 µM dNTP & 2 mM pH 7.5, 10 mM Tris-HCl
& 4 µM primer has Mg 1 mM EDTA
worked with Tag 5'U, the same 50 µg/ml PK
conditions will be used.

used 2 µl of mini-prep - can unknown (hard BMB (3)
still hard to run gel)

Tried dif. enzyme conc 1', 2', 5' and 1:100, 2:100, 5:100
Tag Tag & DV

Colony lysis: Since these colonies were so minute after 10x
at 37° pooled 5 or 6 colonies in a single area -
spotted 2 µl of lysis buffer or H₂O mixed & pipetted out the
liquid on to a tube containing 18 µl of lysis buffer
or H₂O

Colonies in PK lysis 55°, 15' → 95°, 15'
in H₂O 95°, 15'

(Added ~ 5 µl of H₂O) pooled all these tubes together
and made up the volume to 50 µl.

Should have picked more for more reactions
used 10 µl / Rx - appropriately either PK lysed or H₂O lysed ✓
colony itself 10 µl H₂O To Page No. _____

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1/6/95

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For mini-prep DNA : prepared premix with template
 For colony added them later

mini-prep premix : 25x

dNTP 2.5 μ l (200 μ M each / Rx)
 F.P 5 (2.5 μ l) 0.4 μ M
 R.P 5 0.4 μ M
 Template 25x2 \rightarrow last exp. used 1.5 + 1 / Rx
 mini-prep 415 = 2 μ l / Rx
 H₂O

500 \rightarrow 2 μ l / Rx

Premix B: 5x
 (2mM) Buffer B
 100x enzyme 1420
 Tag 1 2 5
 50 50 1 2.5 5
 99.5 99 97.5 95 90 75
 150 \rightarrow 30 μ l / Rx
 Tag + DV
 1:01 2:02 5:08

step 2
 3 cycle

94°, 3'
 20 (94°, 45"
 55°, 30"
 72°, 3')

2 step:

94°, 3'
 94°, 45"
 68°, 5' 20

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age N		2 step		3 step	
Tube #			Tube #		
1		1		13	same as in 2 step.
2		1		14	
3		2		15	
4		2		16	
5		5		17	
6		5		18	
7		1 : .01		19	
8		1 : .01		20	
9		2 : .02		21	
10		2 : .02		22	
11		5 : .05		23	
12		5 : .05		24	

Colonies

In mix A : 15 x

Mix B : 5 x as earlier :
for T + D.V

d.w.TP 15

primer 3

" 3

- 150 (Template 10 µl / Rx like added earlier)

L20 129

150 → 10 µl f Rx

20 µl → + ←

added 30 µl / Rx

appropriately either
Tag alone or Tag + D.V

changing conditions same as mini prep.

for 3 step cycle, 2 step not done. ∴ don't
have much template left from
lysed plasma

Mix # 25 1 + .01

26 2 + .02

27 5 + .05

PK lysed

28 } 11, 20

29 } lysed

30 }

31 }

32 }

33 }

straight pick

35, 36 (20 Tag)

H.w. plain

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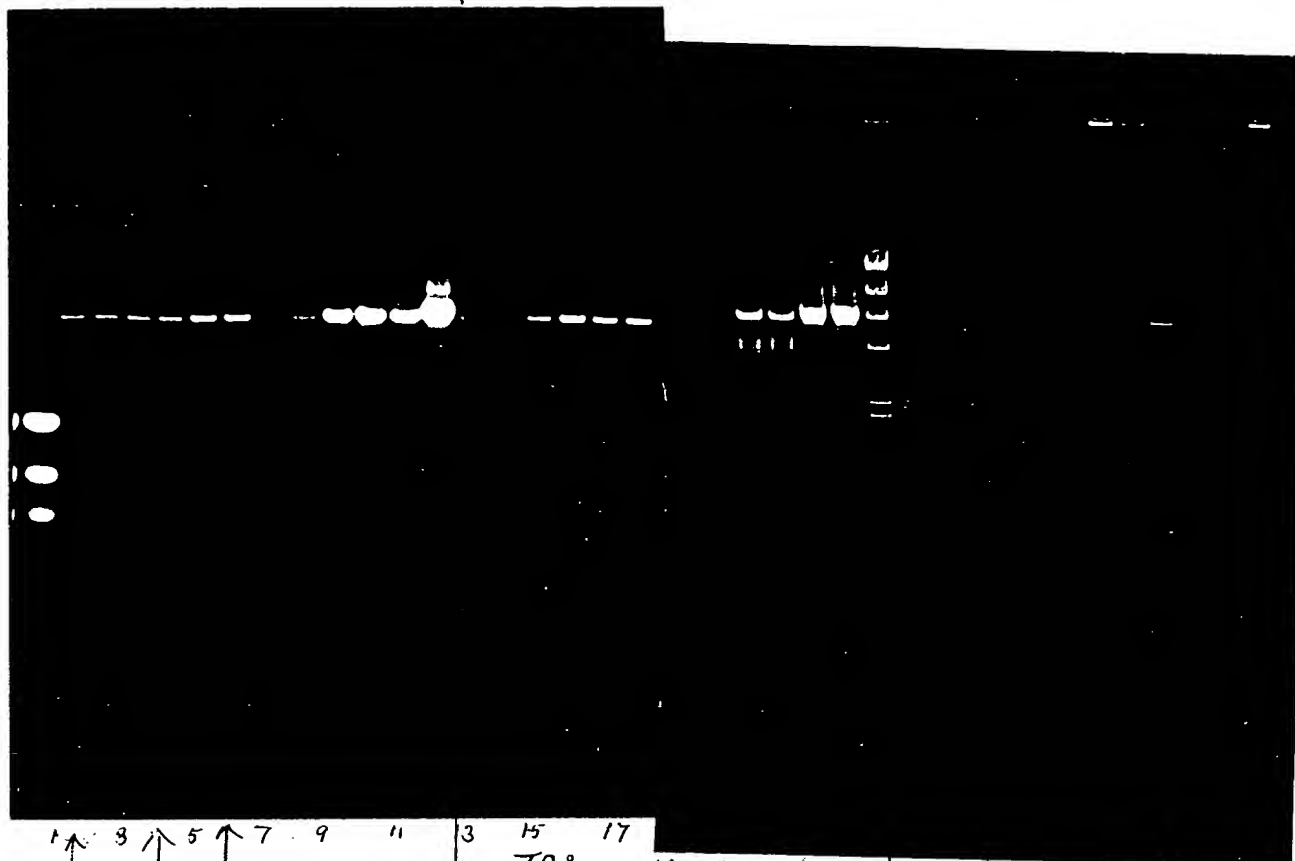
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2 step
cycle

3 step cycle



1 3 5 7 9 11 13 15 17

Tag

19 21 23
T + DVPK H₂O direct peak

Unit 1
enzyme
Tag

2

5

1, 2, 5

Enzyme
Mix

PK H₂O direct peak

W Tag + DV

W Tag + DV

2 U

mini prep.

Plasmids

Result: Even 3 step gave better product with less mis-
plasmid amp should be done under more control.

Witnessed & Understood by me,

Date

1/10/95

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Date

1/9/95

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